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Expression of mammalian cytokines by Trypanosoma cruzi indicat unique signal sequence requirements and processing.

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enabled the expression and secretion of the murine cytokines interleukin-2 (IL-2) a gamma-interferon (gamma-IFN) by transfected T. cruzi. The T. cruzi-derived cytokines were bioactive and produced by both epimastigotes and mammalian forn The native coding sequence of IL-2 was sufficient to cause secretion of the protein the gamma-IFN signal sequence had to be replaced by the IL-2 signal sequence (IL-2/gamma-IFN) to allow efficient secretion of gamma-IFN. The amino acid sequences at the N-termini of the secreted T. cruzi-derived cytokines were differen from the expected murine secreted protein. The secreted IL-2 was cleaved six amir acids downstream from the murine signal sequence cleavage site, and the hybrid IL-2/gamma-IFN molecule was cleaved three amino acids downstream from the predicted signal cleavage site in the IL-2/gamma-IFN molecule. These apparent

differences in signal peptide sequence requirements and cleavage sites most likely indicate that the signal sequence processing in trypanosomes is distinct from that o

A vector based upon the calmodulin-ubiquitin 2.65 locus of Trypanosoma cruzi ha

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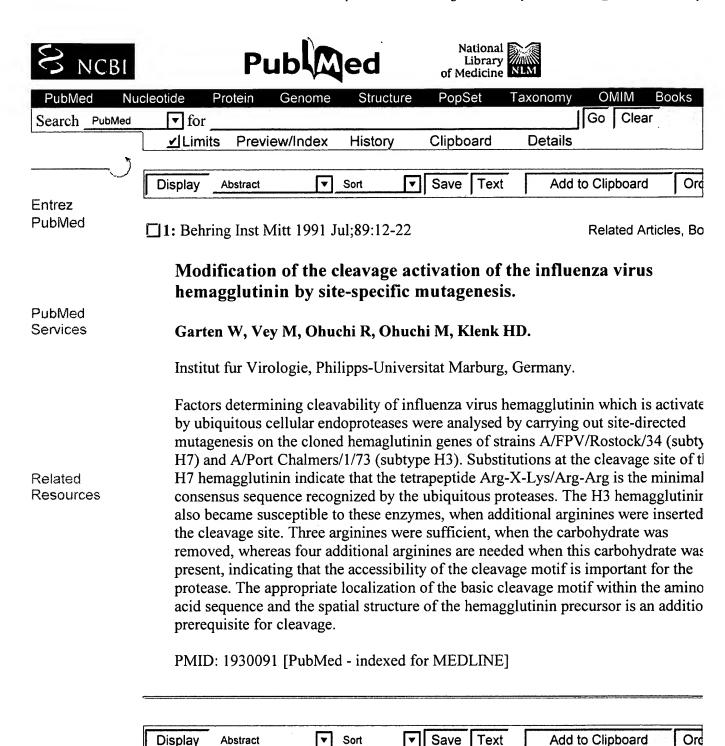
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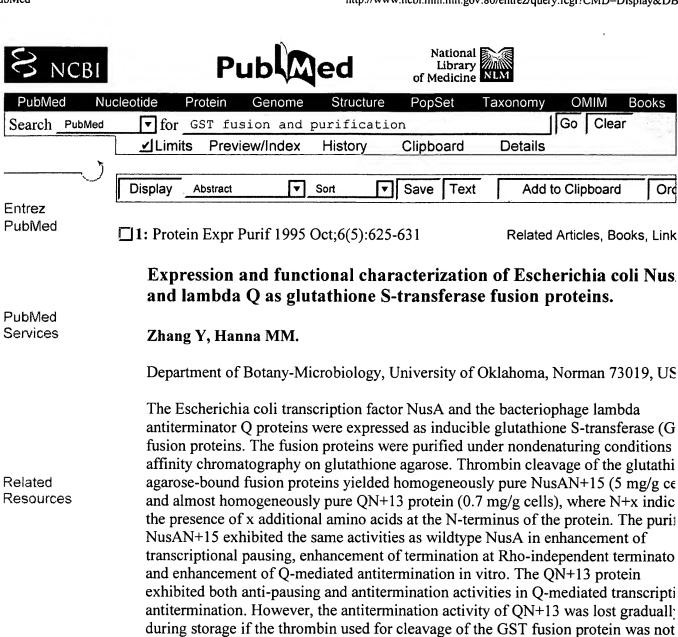


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interaction with RNA polymerase itself.

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removed. This was due to cleavage by thrombin after Arg22 within the Q protein itself, at a noncanonical thrombin cleavage site, so the truncated protein (ON+22) lacked the first 22 amino acids at the N-terminus of Q. The expression vectors described here can be used to rapidly produce large quantities of these proteins, and the truncated Q protein can be used to evaluate the requirement for the N-terminus Q in antitermination, anti-pausing, interactions with the DNA template (qut site), a

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